Substrate influences mating success and transmission of courtship vibrations for the parasitoid *Cotesia marginiventris*

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Abstract

The influences of artificial and natural rearing substrates on mating success were investigated for the parasitoid wasp Cotesia marginiventris (Cresson) (Hymenoptera: Braconidae), a candidate for augmentative biological control of various lepidopteran pests. Five rearing substrates were tested: plastic, glass, chiffon fabric, and leaves of two host plants, bean [Vigna unguiculata (L.) (Fabaceae)] and maize [Zea mays L. (Poaceae)]. Mating success was highest on chiffon, lowest on plastic and glass, and intermediate on maize and bean. The transmission characteristics of one component (buzz 1) of the courtship vibrations produced by male wing fanning were investigated using laser vibrometry. The duration of buzz 1 was longer on maize, bean, and chiffon than on plastic and glass. The fundamental frequency of buzz 1 (~300 Hz) was lowest on bean and highest on glass, and intermediate among other substrates. The relative amplitude of buzz 1 was higher on chiffon than on plastic, glass, or bean, and intermediate on maize. The relative importance of airborne sound and substrate vibration as courtship signals was also investigated with experiments that manipulated the production of courtship vibrations and the mating substrates. The amplitude of courtship vibrations on chiffon was significantly higher for winged males than for dealated males. The mating success of males was impacted by both the presence of wings and the mating substrate. These findings suggest that mating success and transmission of courtship vibrations are influenced by the rearing substrate, and that courtship vibrations are critical to mating success in C. marginiventris. Future efforts to mass rear this parasitoid and other insects should consider the potential influences of rearing substrates on mating.

Introduction

Insect mating is often influenced by chemical and physical environmental factors, such as host plant odors, ambient temperatures, and time of day (van den Assem, 1986; Quicke, 1997), but the influence of the mating substrate and its physical properties on mating success has rarely been investigated. Several studies found that the mating success of some insects and other arthropods that use vibrational communication during courtship can be affected by the mating substrate (Miklas et al., 2001; Elias et al., 2004), while others found that the physical properties of the substrate had little influence on the transmission of courtship vibrations (Henry & Martinez Wells, 2004; Cocroft et al., 2006).

Male parasitoid wasps from several families of Hymenoptera, including those of *Cotesia* spp. Cameron (Hymenoptera: Braconidae), fan their wings during courtship, producing species-specific, low amplitude sounds, and substrate vibrations, which are likely to play an important role in courtship communication (Vinson, 1972; Leonard & Ringo, 1978; van den Assem & Putters, 1980; Sivinski & Webb, 1989; Field & Keller, 1993; Kimani & Overholt, 1995). Transmission of courtship vibrations in the substrate appeared critical for mating in the parasitoid

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Cotesia rubecula (Marshall) (Field & Keller, 1993). Also, replay of courtship sounds to *Nasonia vitripennis* (Walker) (Hymenoptera: Pteromalidae) increased mating for older males, suggesting that courtship acoustics convey information concerning male quality (van den Assem & Putters, 1980). The transmission of parasitoid courtship vibrations may be influenced by host plant substrates and artificial substrates used for mass rearing in the laboratory, which in turn could influence their mating success.

Transmission of courtship vibrations through substrates used in insect mass-rearing programs has not been studied. Courtship vibrations may transmit differently in materials commonly associated with rearing, such as plastic, glass, and fabric, than in natural host plant substrates. A common goal in parasitoid mass rearing for biological control is to produce as many females as possible, because only females attack and kill hosts in the field. Most parasitoid wasps reproduce by arrhenotoky, thus inseminated females produce female and male offspring, while virgin females produce only males. Selecting rearing substrates that facilitate courtship communication and mating could increase the number of female parasitoids produced under mass rearing.

Cotesia marginiventris (Cresson) is a parasitoid wasp whose hosts are moth larvae (Lepidoptera: Noctuidae) and is an important natural control agent of several pest species, including Spodoptera frugiperda (J. E. Smith) and Trichoplusia ni (Hübner) (McCutcheon & Turnipseed, 1981; Braman & Yeargan, 1991; Novoa & Luna, 1996; Gillespie et al., 1997). Cotesia marginiventris is a solitary endoparasitoid that attacks first and second instars (Boling & Pitre, 1970; Kunnalaca & Mueller, 1979; Tillman, 2001) and has been considered for augmentative biological control of T. ni in greenhouses (Gillespie et al., 1997; Urbaneja et al., 2002; Riddick, 2006). The sex ratios (% males) of C. marginiventris recorded under mass rearing and field conditions vary between 40 and 80% (Jalali et al., 1987; Riggin et al., 1992; Novoa & Luna, 1996; Gillespie et al., 1997; Tillman, 2001). The male-biased sex ratios observed in some studies may have been due to a high frequency of virgin females, although the potential role of mating substrates and transmission of courtship vibrations in rearing materials were not investigated.

The objective of this study was to assess the influence of natural and artificial rearing substrates on the mating success and transmission of courtship vibrations for the parasitoid *C. marginiventris*. In addition, the importance of airborne sound relative to substrate vibrations produced during courtship was determined by experimentally manipulating the production of courtship vibrations on two substrates and quantifying their influence on mating success.

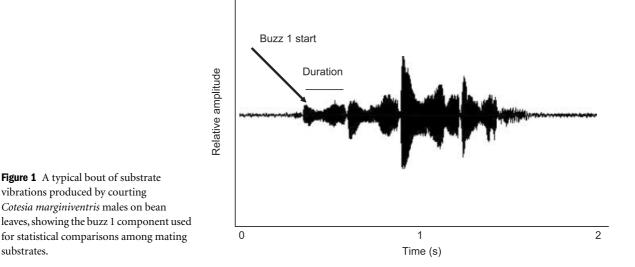
Materials and methods

Insects

A culture of C. marginiventris was maintained on larvae of the moth S. frugiperda. Parasitoid pupae were originally obtained from a laboratory colony in Tifton, GA, USA. Spodoptera frugiperda adults were maintained in 2.4-l white plastic containers with 20% (wt/vol) sucrose solution as food, at 27 ± 2 °C, $60 \pm 5\%$ r.h., and a photoperiod of L14:D10. Adult moths oviposited on a paper towel, and emerging S. frugiperda larvae were fed with a maize earworm diet (Bio-serv Inc., Frenchtown, NJ, USA). To produce parasitoids, 20 S. frugiperda second instars, larval diet, and a streak of honey were placed into a 24-ml glass vial with one adult male and female C. marginiventris (0-48 h old) for 48 h. Larvae were then transferred to plastic cups with diet until parasitoids formed cocoons. Each C. marginiventris pupa was then isolated in a 1-ml glass vial (30 × 8 mm; Bioquip, Gardena, CA, USA) so that emerging adults would remain virgin until used in an experiment. All adult C. marginiventris were stored overnight at 15 °C before use in mating trials, to increase the likelihood of mating (A Joyce, unpubl.).

Mating success on five substrates

The experiment was designed to assess the effect of rearing materials on the mating success of individual C. marginiventris females, and consisted of five treatments: glass shell vials $(1 \text{ ml}, 30 \times 8 \text{ mm}; \text{Bioquip})$, plastic Petri dishes $(50 \times 9 \text{ mm},$ Falcon®; Becton Dickinson Labware, Franklin Lakes, NJ, USA), white chiffon fabric (Hancock Fabrics; Chiffon Georgette, 100% polyester), and two host plant substrates: maize leaves [Zea mays L (Poaceae), Pioneer® seed 34A55; Johnston, IA, USA] and bean leaves [Vigna unguiculata (L.) (Fabaceae), California blackeye cowpea no. 5; Gurney's Seed and Nursery Co., Greendale, IN, USA]. Glass shell vials were plugged with cotton to prevent parasitoids from escaping. The chiffon, bean leaf, and maize leaf treatments also consisted of a plastic component, to enclose the insects on the substrate. A chiffon fabric arena was constructed of a plastic Petri dish $(50 \times 9 \text{ mm})$ with a circular opening (40 mm diam) cut in the lid, with chiffon fabric glued over the opening, such that the surface was smooth and taut. The maize and bean leaf arenas consisted of a leaf, which remained attached to the plant, covered with a plastic vial (160 ml, 85×50 mm; Bioquip). The five rearing materials varied from thickest to thinnest in the following order: plastic, glass, bean leaf, maize leaf, and chiffon fabric. Although the construction of arenas for leaves and the chiffon material required plastic support material, all matings occurred on the substrate associated with that treatment.



Each male–female pair of virgin parasitoids were monitored for 10 min on each of the five treatments at laboratory temperature and humidity ($25 \pm 2 \degree C$, 50% r.h.), between 08:00 and 10:00 hours, and mating success or failure was recorded. All parasitoids used in this and subsequent experiments were 2–4 days old, and for this experiment, 34 pairs were observed per substrate. Each experimental arena and all parasitoids were used only once and then discarded. A χ^2 goodness-of-fit test was used to determine whether mating frequencies differed among the five treatments. Post-hoc separation of mating frequencies on each treatment was by a modified Tukey test for proportions (Zar, 1999).

Transmission of courtship vibrations on five substrates

Courtship vibrations produced by male wing fanning were recorded on the five substrates described above to determine whether the substrates influenced their transmission. Male courtship primarily consists of wing fanning when in close proximity (1–2 cm) to a female. All parasitoids were used for only one recording. Males were recorded courting a female on each substrate for 10 min. All recordings were made under standardized laboratory conditions ($25 \pm 2 \,^{\circ}$ C, 50% r.h.) using a randomized block design. Each block consisted of a single replicate (male–female pair) on each of the five substrates and the order of substrates was randomized within each block. Ten pairs were recorded courting on each of the five substrates.

Recordings were made with a laser Doppler vibrometer (Model OFV 353; Polytec Inc., Irvine, CA, USA) at 1 mm s⁻¹ volt⁻¹ sensitivity in an enclosed, sound reduction chamber (~1 m long × 1 m high × 0.75 m wide), located on a vibration isolation table (TMCTM, Model NAF 2000;

Peabody, MA, USA). One male–female pair was placed on one of the substrates. The laser was positioned above the test substrate, and the laser beam was focused on reflective tape (4 mm²) on the test surface of the substrate of interest. The vibrational signal detected by the laser was digitized and recorded on a Macintosh® computer equipped with an Audiomedia III sound card (16 bit, sampling rate 44.1 kHz). Peak® software (version 3.0; Bias, Petaluma, CA, USA) was used to record the signals. The distance between the laser beam point of contact on the substrate and the location of the courting male was monitored and recorded in 1 cm increments. Only courtship vibrations produced by males within 2 cm of the laser were used for analyses.

Vibrations produced by courting C. marginiventris males consist of several long buzzes, which may be preceded and/ or followed by shorter pulses (Figure 1). Several parameters of the first buzz component (hereafter buzz 1) were used for comparison of vibration transmission among the five substrates (Figure 1). Cool Edit Pro software (version 2.0, now Adobe Audition 2.0; San Jose, CA, USA) was used to measure the duration, frequency, and relative amplitude of the buzz 1. The buzz 1 duration was measured in ms. The fundamental frequency (Hz) and its relative amplitude (-db, reference 0 db) were measured 10 ms after the beginning of buzz 1. Analyses were based on relative amplitudes rather than absolute amplitudes (i.e., velocity), because it was not practical to calibrate the absolute amplitude of each measurement while maintaining parasitoid age constant for recordings.

Statistical comparison of the fundamental frequency and the relative amplitude among the treatments was done by a one-way analysis of variance (ANOVA), and Tukey's post-hoc test was used to separate means (SPSS, 2001). A non-parametric Kruskal–Wallis test was used to compare the buzz 1 duration among treatments (Statistix, 2000), with post-hoc comparisons of means as described in Siegel & Castellan (1988), as data did not meet the assumptions of ANOVA.

Role of airborne sound and substrate vibration in mating

Two experiments were conducted to assess how mating success is influenced by airborne sounds relative to substrate vibrations. The first experiment assessed whether wing ablation affected substrate vibrations produced by courting males. This was done by quantitative comparison of the peak velocity (mm s⁻¹) of substrate vibrations produced by courting normal and dealated males on the chiffon arena. This was a calibrated amplitude measurement, which differed from the relative amplitude measurement in the section above. A calibration procedure was performed prior to each recording, following procedures available in SpectraPro (version 3.32c; Sound Technology, Inc., El Cajon, CA, USA). A 200 Hz sine wave was output from a PC to a Tascam (Montello, CA, USA) DA-3-MP digital tape recorder, which served as an amplifier. The output from the tape recorder headphone jack was fed to a Brüel & Kjaer (Naerum, Denmark) Model 4810 minishaker positioned at the center of the arena. The volume was adjusted to within the range typical of wasp signals (A Joyce, unpubl.). Simultaneously, the output of the laser decoder was sent to SpectraPro which detected relative voltage. Actual output voltage was measured directly using a Fluke® 8060A multimeter (Fluke, WA, USA), and was then entered and used by SpectraPro to calibrate subsequent measurements.

Both dealated and normal males were prepared for recording trials by chilling in vials in a freezer for 10 min, and then removing them to a frozen cold pack. Each male was then placed dorsal side up under a dissecting microscope, and the thorax was pressed gently with forceps to spread the wings. Dealated males were prepared by ablating each wing near the base, leaving approximately 1/4 of each wing. Normal males were mock ablated using forceps to touch the wings but not ablate them.

Courting dealated males were recorded on a chiffon arena prior to wing ablation, and then again after wing ablation, to determine the magnitude of any change in courtship vibration amplitude. Normal courting males were similarly recorded prior to and after mock wing ablation. The order of recordings of normal and dealated males was alternated. The first courting bout for each male was used to measure peak velocity of the courtship vibration. Paired t-tests were used to compare the peak velocities for dealated males prior to and after wing ablation, and for normal males prior to and after mock wing ablation. Ten normal males and 10 dealated males were recorded for comparisons. The second experiment compared the mating success of normal and dealated males on a good (chiffon) and a poor (glass) artificial mating substrate, according to the mating success and transmission characteristics shown for those substrates (see Results). The expectation was that if courtship signaling relied exclusively on airborne (near-field) sound, then wing ablation, but not substrate, would affect mating success. In contrast, if courtship signaling relied exclusively on substrate vibrations, then substrate and not wing ablation would affect the mating success of males.

Each male, normal or dealated, was prepared for trials as described above and was then paired individually with a female for up to 10 min and mating success or failure was recorded. Eighteen normal and 18 dealated males were tested on both chiffon and on glass substrates, with the order of substrates alternated. Logistic regression was used to assess the relative importance of wings and substrate to mating success (Stata, 2005). Mating success was the dependent variable, and substrate (glass = 0, chiffon = 1) and wings (dealated = 0, winged = 1) were independent predictor variables.

Results

Mating success on five substrates

The mating success of females differed significantly among treatments ($\chi^2 = 17.74$, d.f. = 4, and P = 0.001). Mating success was greatest on chiffon, intermediate on maize and bean, and lowest on glass and plastic (Figure 2).

Transmission of courtship vibrations on five substrates

The measurable mean duration of buzz 1 varied significantly among treatments (Kruskal–Wallis: $\chi^2 = 23.22$, d.f. = 4, and P<0.001). The mean duration on plastic was significantly

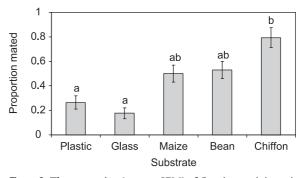


Figure 2 The proportion (mean \pm SEM) of *Cotesia marginiventris* females successfully mating on each of five substrates (n = 34 pairs/substrate). Different letters indicate significant differences (P<0.05) (χ^2 test, followed by a Tukey-type comparison for proportions).

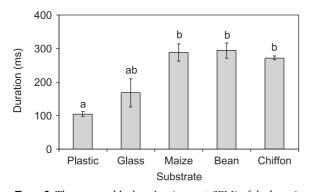


Figure 3 The measurable duration (mean \pm SEM) of the buzz 1 component of courtship produced by male *Cotesia marginiventris* on each of five substrates (n = 10 males/ substrate). Different letters indicate significant differences (P<0.05) (Kruskal–Wallis test, followed by pairwise comparisons; Siegel & Castellan, 1988).

shorter than on maize, bean, and chiffon, and on glass it was intermediate between plastic and the other substrates (Figure 3).

The mean fundamental frequencies at the beginning of buzz 1 for males on the five treatments ranged from 295 to 319 Hz, and they differed significantly among substrates (ANOVA: $F_{4,45} = 16.24$, P = 0.04; Figure 4). Bean had a significantly lower mean frequency than glass, while the fundamental frequencies on other substrates were intermediate and not significantly different from each other (Figure 4). The mean relative amplitude of the fundamental frequency varied significantly among treatments (ANOVA: $F_{4,45} = 7.43$, P<0.001), and was significantly higher on chiffon than on plastic, glass, and bean, and intermediate on maize (Figure 5).

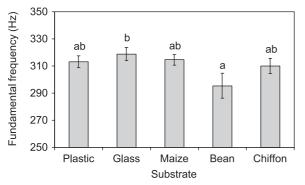


Figure 4 The fundamental frequency (mean \pm SEM) at the beginning of buzz 1 (n = 10 males/substrate). Different letters indicate significant differences (P<0.05) [one-way analysis of variance (ANOVA), followed by Tukey's test].

Role of airborne sound and substrate vibration in mating

Both dealated and normal males courted females on chiffon arenas and produced detectable substrate vibrations (Figure 6). The mean peak velocity of courtship vibrations produced by dealated males was reduced by 72% after wing ablation (t = 3.86, d.f. = 8, and P = 0.005), while no change was detected in normal males after mock wing ablation (t = 0.02, d.f. = 7, and P = 0.988; Figure 6).

Both the mating substrate and the presence of wings significantly predicted the probability of mating (Table 1). The odds of mating were 4.3 times greater on chiffon than on glass (P = 0.01), and 9.6 times greater for normal winged males than for dealated males (P<0.001; Table 1). Normal males mated more frequently than dealated males on both substrates, with a higher proportion of normal and dealated males mating on chiffon than on glass, and no mating was observed for dealated males on glass (Figure 7).

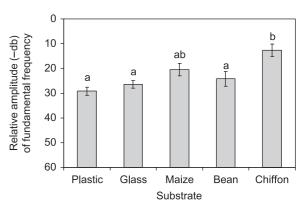


Figure 5 The relative amplitude (mean \pm SEM) of the fundamental frequency at the beginning of buzz 1 (n = 10 males/ substrate). Different letters indicate significant differences (P<0.05) [one-way analysis of variance (ANOVA), followed by Tukey's test].

Discussion

Mating success on five substrates

The mating success of *C. marginiventris* was highest on chiffon, an artificial rearing substrate, followed by the two host plants, bean and maize, and lowest on glass and plastic (Figure 2). Greater mating frequencies on host plants than on artificial substrates might be expected, as plant volatiles are important cues in long range attraction and mate location (McAuslane et al., 1990). However, little is known about how plants and artificial substrates affect close range courtship communication and mating in parasitoids. That

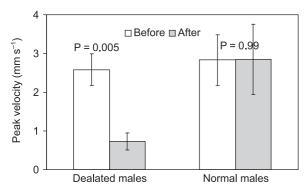


Figure 6 The peak velocity (mean \pm SEM) on chiffon arenas from the first courting bout of male *Cotesia marginiventris*, for dealated males before and after wing ablation, and for normal males before and after mock ablation. P-values correspond to paired t-tests (n = 8/dealated males, n = 7/normal males).

mating success was highest on chiffon suggests that physical properties of the substrate are important for mating in *C. marginiventris*. Prior studies showed that vibrations transmit differently through common building materials (Rossing & Fletcher, 2004), and that physical properties of the substrate may affect vibrational signals and mating success in arthropods (Miklas et al., 2001; Elias et al., 2004).

Parasitoid sex ratios that are male biased in laboratory cultures could be partially attributable to a high frequency of virgin parental females. For example, *C. marginiventris* sex ratios were more male biased in several laboratory studies relative to field studies (Novoa & Luna, 1996; Gillespie et al., 1997). *Cotesia melanoscela* (Ratzeburg), a parasitoid of the gypsy moth, had 65% males in massrearing programs in which acrylic plastic mating cages were used (Kolodny-Hirsch, 1988), while another rearing program for *C. melanoscela* that used mating cages constructed of paper and cloth, and contained plants, produced 50% males (Webb et al., 1997). In these studies, it is plausible that a high frequency of virgin parental females and consequently male-biased sex ratios could be due to the absence of an appropriate mating substrate.

Table 1 Logistic regression for probability of mating forCotesia marginiventris with two predictor variables, substrate(glass, chiffon) and wings (normal, dealated) (n = 72)

Predictor	β	Odds	Wald's test	Р
variable	coefficient	ratio	Z	
Substrate	1.45	4.29	2.4	0.010
Wings	2.26	9.63	3.56	<0.001

Transmission of courtship vibrations on five substrates

The measurable mean duration of buzz 1 was longer on maize, bean, and chiffon than on plastic, and was intermediate on glass (Figure 3). The durations on maize, bean, and chiffon were similar to the duration of buzz 1 found in a previous study of C. marginiventris (Sivinski & Webb, 1989). All C. marginiventris males appeared to court similarly on the five substrates tested, although the possibility cannot be eliminated that their courtship behavior changed on plastic or glass. The shorter mean duration of buzz 1 on plastic and glass may have been an underestimate, because its lower amplitude on these substrates was often near the lower detection threshold of the laser vibrometer. Similarly, due to the shorter duration of the courtship vibration on those substrates, female C. marginiventris may not detect the entire length of buzz 1 on plastic or glass, leading to reduced mating success. This is consistent with the findings that mating success of C. marginiventris was higher on the substrates with longer buzz 1 durations. In a study of the stinkbug Nezara viridula L. (Heteroptera: Pentatomidae), longer artificial courtship

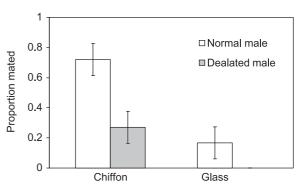


Figure 7 The proportion (mean \pm SEM) of normal and dealated *Cotesia marginiventris* males mating on two substrates, glass and chiffon (n = 18 pairs/column). See Table 1 for logistic regression statistics.

signals (600–800 ms) played to males elicited stronger responses relative to shorter signals (<600 ms) (Miklas et al., 2001). Also, species recognition is often influenced by the duration of courtship sounds and vibrations (Greenfield, 2002). For example, *Chrysoperla carnea* (Stephens) (Neuroptera: Chrysopidae) lacewings respond more frequently to conspecific relative to heterospecific courtship vibrations, which vary in duration and frequency (Henry et al., 2002).

The mean frequency of courtship vibrations at the beginning of buzz 1 was significantly lower on bean than

on glass (Figure 4). Courtship vibration frequencies may transmit differently through artificial substrates, such as plastic or glass, relative to leaves of host plants, such as maize or bean. Prior studies have shown that leaves can filter signals (Casas & Magal, 2006), allowing transmission of lower frequencies while filtering out higher frequencies. In particular, leaf veins attenuate signals more than leaf lamina (Magal et al., 2000). In our study, the bean leaves had substantially more leaf veins than the maize leaves, which may have contributed to frequency filtering on bean.

The high relative amplitude on chiffon may have been from chiffon being the thinnest substrate evaluated. As noted by others, thicker substrates more than thinner ones can attenuate the amplitude of vibratory signals (Fischer et al., 2003; Cocroft et al., 2006). In this study, *C. marginiventris* mated most frequently on chiffon. Higher courtship vibration amplitudes on some substrates, such as chiffon, may reach a minimum threshold needed to stimulate vibratory receptors sooner (Čokl et al., 2006), and thus be detected more rapidly by females.

Few differences were evident in signal-transmission characteristics between the two host plants that were evaluated. There was no difference in the duration of buzz 1 on bean or corn (Figure 3), and relative amplitudes of the fundamental frequency measured at 0-2 cm from the courting male were similar in both host plants (Figure 5). Cocroft et al. (2006) recorded courtship vibrations of the treehopper Umbonia crassicornis (Amyot and Serville) (Homoptera: Membracidae) on a woody host and a woody non-host plant, and did not find significant differences in signal parameters between the two plants at close range (5 cm). However, signal transmission was affected by distance, and by 15 cm there were significant differences in the final frequency and the signal duration. Similarly, few differences in signal transmission between host and non-host plants were observed for courtship vibrations of the lacewings Chrysoperla downesi (Smith) and Chrysoperla plorabunda (Fitch), although signal amplitude attenuated by a distance of 20 cm (Henry & Martinez Wells, 2004).

Other studies involving arthropods show that substrates influence the transmission of vibrational signals associated with foraging. Substrate density influences the ability of two ichneumonid wasps, *Pimpla turionellae* L. and *Xanthopimpla stemmator* (Thunberg), to locate their hosts (Fischer et al., 2003), perhaps due to the effect of substrate density on signal amplitudes. Substrates also influence the transmission of honeybee waggle dance vibrations. Transmission of artificial vibrations of 250 Hz had greater amplitude in honeybee combs made of beeswax than in artificial plastic combs (Seeley et al., 2005).

Role of airborne sound and substrate vibration in mating

The velocity of courtship vibrations produced by *C.* marginiventris males was within the range of vibrations produced and detected by other species of courting insects. Vibrational courtship signals with velocities as low as 0.1 mm s⁻¹ are considered in the range of detection for insects (Michelsen et al., 1982). Courting male *C. marginiventris* produced vibrations with a mean peak velocity of 2.58 mm s⁻¹ on chiffon fabric (Figure 6), which is comparable to values of vibrational courtship signals from *N. viridula* males, which vary between 2 and 3.5 mm s⁻¹ (Miklas et al., 2001).

Both wings and substrate were important to mating success (Figure 7; Table 1), which suggests that female C. marginiventris respond to both substrate vibrations and near-field sound, both detectable by Hymenoptera (Towne & Kirchner, 1989; Meyhöfer & Casas, 1999; Tomov et al., 2003). Some studies that compared the importance of the near-field sound and substrate vibrations potentially used by parasitoids as courtship signals found that mating success was lower in dealated parasitoid males (Miller & Tsao, 1974; Kitano, 1975; Rungrojwanich & Walter, 2000), but the presence of wings appeared more important for some species than others (van den Assem & Putters, 1980; Ruther et al., 2000). A prior study showed that substrate vibrations produced during courtship are essential for mating in C. rubecula (Field & Keller, 1993). Vibrational signaling during courtship appears important as well for C. marginiventris, and may be used by other species of parasitoids, particularly in the genus Cotesia and the family Braconidae (Leonard & Ringo, 1978; van den Assem & Putters, 1980; Sivinski & Webb, 1989).

Courtship signals other than vibrations, such as visual and chemical cues, may have been used by C. marginiventris, but it is unlikely that the substrates influenced those signals and impacted female mating success. Chemical cues, such as pheromones, would likely be similar among substrates, although their production or effect may be influenced by the presence of a plant odor (McAuslane et al., 1990). Nevertheless, mating success on chiffon was similar to that on host plants, and significantly higher than on plastic or glass, indicating that physical properties of the substrate and/or physical cues such as courtship vibrations are important for mating success in this parasitoid. As other orders of insects are known to use vibrational communication (Cocroft & Rodriguez, 2005), knowledge of the transmission characteristics of their courtship signals through substrates could be important for efficiently mass rearing them as well.

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References

- van den Assem J (1986) Mating behaviour in parasitic wasps. Insect Parasitoids (ed. by J Waage & D Greathead), pp. 137– 167. Academic Press, London, UK.
- van den Assem J & Putters FA (1980) Patterns of sound produced by courting chalcidoid males and its biological significance. Entomologia Experimentalis et Applicata 27: 293–302.
- Boling JC & Pitre HN (1970) Life history of *Apanteles marginiventris* with descriptions of immature stages. Journal of the Kansas Entomology Society 43: 465–470.
- Braman SK & Yeargan KV (1991) Reproductive strategies of primary parasitoids of the green cloverworm (Lepidoptera: Noctuidae). Environmental Entomology 20: 349–353.
- Casas J & Magal C (2006) Mutual eavesdropping through vibrations in a host-parasitoid interaction: from plant biomechanics to behavioural ecology. Insect Sounds and Communication: Physiology, Behaviour, Ecology and Evolution (ed. by S Drosopoulos & MF Claridge), pp. 263–271. CRC, Boca Raton, FL, USA.
- Cocroft RB & Rodriguez RL (2005) The behavioral ecology of insect vibrational communication. Bioscience 55: 323–334.
- Cocroft RB, Shugart HJ, Konrad KT & Tibbs K (2006) Variation in plant substrates and its consequences for insect vibrational communication. Ethology 112: 779–789.
- Čokl A, Virant-Doberlet M & Zorović M (2006) Sense organs involved in the vibratory communication of bugs. Insect Sounds and Communication: Physiology, Behaviour, Ecology and Evolution (ed. by S Drosopoulos & MF Claridge), pp. 71– 80. CRC, Boca Raton, FL, USA.
- Elias DO, Mason AC & Hoy RR (2004) The effect of substrate on the efficacy of seismic courtship signal transmission in the jumping spider *Habronattus dossenus* (Araneae: Salticidae). Journal of Experimental Biology 207: 4105–4110.
- Field SA & Keller MA (1993) Courtship and intersexual signaling in the parasitic wasp *Cotesia rubecula* (Hymenoptera: Braconidae). Journal of Insect Behavior 6: 737–750.
- Fischer S, Samietz J & Dorn S (2003) Efficiency of vibrational sounding in parasitoid host location depends on substrate density. Journal of Comparative Physiology A 189: 723–730.
- Gillespie D, Opit G, McGregor R, Johnston M, Quiring D & Foisy M (1997) Recommendations for the use of the parasitic wasp, *Cotesia marginiventris* (Hymenoptera: Braconidae) for biological control of cabbage looper, *Trichoplusia ni*. Pacific Agri-Food Research Centre (Agassiz) Technical Report 142: 1–2.
- Greenfield MD (2002) Signalers and Receivers: Mechanisms and Evolution of Arthropod Communication. Oxford Press, Oxford, UK.

- Henry CS, Brooks SJ, Duelli P & Johnson JB (2002) Discovering the true *Chrysoperla carnea* (Insecta: Neuroptera: Chrysopidae) using song analysis, morphology and ecology. Annals of the Entomological Society of America 95: 172–191.
- Henry CS & Martinez Wells ML (2004) Adaptation or random change? The evolutionary response of songs to substrate properties in lacewings (Neuroptera: Chrysopidae: Chrysoperla). Animal Behaviour 68: 879–895.
- Jalali SK, Singh SP & Ballal CR (1987) Studies on host age preference and biology of the exotic parasite, *Cotesia marginiventris* (Cresson) (Hymenoptera: Braconidae). Entomophaga 12: 59–62.
- Kimani SW & Overholt WA (1995) Biosystematics of the *Cotesia flavipes* complex (Hymenoptera: Braconidae): interspecific hybridization, sex pheromone and mating behaviour studies. Bulletin of Entomological Research 85: 379–386.
- Kitano H (1975) Studies on the courtship behavior of *Apanteles* glomeratus L. 2. Role of the male wing during courtship and the releaser of mounting and copulatory behavior in the males. Kontyu 43: 513–521.
- Kolodny-Hirsch DM (1988) Influence of some environmental factors on the laboratory production of *Cotesia melanoscela* (Braconidae: Hymenoptera): a larval parasitoid of *Lymantria dispar*. Environmental Entomology 17: 127–131.
- Kunnalaca S & Mueller AJ (1979) A laboratory study of *Apanteles marginiventris*, a parasite of green cloverworm. Environmental Entomology 8: 365–368.
- Leonard SH & Ringo JM (1978) Analysis of male courtship patterns and mating behavior of *Brachymeria intermedia*. Annals of the Entomological Society of America 71: 817–826.
- Magal C, Schöller M, Tautz J & Casas J (2000) The role of leaf structure in vibration propagation. Journal of the Acoustical Society of America 108: 2412–2418.
- McAuslane HJ, Vinson SB & Williams HJ (1990) Influence of host plant on mate location by the parasitoid *Campoletis sonorensis* (Hymenoptera: Ichneumonidae). Environmental Entomology 19: 26–31.
- McCutcheon GS & Turnipseed GS (1981) Parasites of lepidopterous larvae in insect resistant and susceptible soybeans in South Carolina. Environmental Entomology 10: 69–74.
- Meyhöfer R & Casas J (1999) Vibratory stimuli in host location by parasitic wasps. Journal of Insect Physiology 45: 967–971.
- Michelsen A, Fink F, Gogala M & Traue D (1982) Plants as transmission channels for insect vibrational songs. Behavioral Ecology and Sociobiology 11: 269–281.
- Miklas N, Stritih N, Čokl A & Virant-Doberlet M (2001) The influence of substrate on male responsiveness to the female calling song in *Nezara viridula*. Journal of Insect Behavior 14: 313–332.
- Miller MC & Tsao CH (1974) Significance of wing vibration in male *Nasonia vitripennis* (Hymenoptera: Pteromalidae) during courtship. Annals of the Entomological Society of America 67: 772–774.
- Novoa MC & Luna MG (1996) Parasitism, survivorship, sex ratio, and developmental time of *Cotesia marginiventris* (Cresson) (Hymenoptera: Braconidae), parasitizing *Rachiplusianu* (Guennée) (Lepidoptera: Noctuidae) larvae in soybean crops in Argentina. Acta Entomologica Chilena 20: 23–28.

- Quicke DLJ (1997) Parasitic Wasps. Chapman & Hall, London, UK.
- Riddick EW (2006) Egg load and body size of lab cultured *Cotesia* marginiventris. BioControl 51: 603–610.
- Riggin TM, Isenhour DJ & Espelie KE (1992) Effect of *Cotesia marginiventris* (Hymenoptera: Braconidae) when rearing host fall armyworm (Lepidoptera: Noctuidae) on meridic diet containing foliage from resistant or susceptible corn genotypes. Environmental Entomology 21: 214–219.
- Rossing TD & Fletcher NH (2004) Principles of Vibration and Sound, 2nd edn. Springer, New York, NY, USA.
- Rungrojwanich K & Walter GH (2000) The Australian fruit fly parasitoid *Diachasmimorpha kraussii* (Fullaway): mating behavior, modes of sexual communication and crossing tests with *D. longicaudata* (Ashmead) (Hymenoptera: Braconidae: Opiinae). Pan-Pacific Entomologist 76: 12–23.
- Ruther JM, Homann M & Steidle JLM (2000) Female-derived sex pheromone mediates courtship behaviour in the parasitoid *Lariophagus distinguendus*. Entomologia Experimentalis et Applicata 96: 265–274.
- Seeley TD, Reich AM & Tautz J (2005) Does plastic comb foundation hinder waggle dance communication? Apidologia 36: 513–521.
- Siegel S & Castellan NJ Jr (1988) Nonparametric Statistics for the Behavioral Sciences, 2nd edn. McGraw-Hill, New York, NY, USA.
- Sivinski J & Webb JC (1989) Acoustic signals produced during courtship in *Diachasmimorpha* (= *Biosteres*) *longicaudata* (Hymenoptera: Braconidae) and other Braconidae. Annals of the Entomological Society of America 82: 116–120.

- SPSS (2001) SPSS Syntax Reference Guide. SPSS Inc, Chicago, IL, USA.
- Stata (2005) Base Reference Manual. Stata, College Station, TX, USA.
- Statistix (2000) Statistix 7 User's Manual. Analytical Software, Tallahase, FL, USA.
- Tillman P (2001) Factors affecting parasitization of *Spodoptera exigua* (Lepidoptera: Noctuidae) and sex ratio of the parasitoid *Cotesia marginiventris* (Hymenoptera: Braconidae). Journal of Entomological Science 36: 188–198.
- Tomov BW, Bernal JS & Vinson SB (2003) Impacts of transgenic sugarcane expressing GNA Lectin on parasitism of Mexican Rice Borer by *Parallorhogas pyralophagus* (Marsh) (Hymenoptera: Braconidae). Environmental Entomology 32: 866–872.
- Towne WF & Kirchner WH (1989) Hearing in honey bees: detection of air-particle oscillations. Science 244: 686–688.
- Urbaneja A, van der Blom J, Lara L, Timmer R & Blockmans R (2002) Utilización de *Cotesia marginiventris* (Cresson) (Hym.: Braconidae) para el control biológico de orugas (Lep.: Noctuidae) en el manejo integrado de plagas en pimiento bajo invernadero. Boletin Sanidad Vegetal y Plagas 28: 239–250.
- Vinson SB (1972) Courtship behavior and evidence for a sex pheromone in the parasitoid *Campoletis sonorensis* (Hymenoptera: Ichneumonidae). Environmental Entomology 1: 409–414.
- Webb RE, Venables L & Thorpe KW (1997) Improved rearing and release procedures for *Cotesia melanoscela* (Hym: Braconidae), an early season parasitoid of *Lymantria dispar* (Lep: Lymantriidae). Entomophaga 42: 471–481.
- Zar JH (1999) Biostatistical Analysis, 4th edn. Prentice Hall, Upper Saddle River, NJ, USA.